

in a free standing silicon nitride membrane. The translocation of CTPR proteins was measured in KCl solution at pH below and above its isoelectric point (pI), as well as with and without denaturing agent, Guanidine HCl. When a CTPR protein molecule transits through a nanopore driven by an applied voltage, it partially blocks the ions (K^+ and Cl^-) flow in the nanopore and generates a characteristic electric current blockage signal. The current blockage signal reveals information about the size, conformation, and primary sequence of the CTPR protein molecule. Previous translocation studies carried out with DNA have established that higher bias voltages result in shorter duration current blockages indicating that DNA translocates faster at a stronger electric field. However, CTPR translocation studies presented here show that longer duration current blockades were observed at higher bias voltages. We explain this surprising result by theoretical analysis of CTPR protein translocation in solid state nanopores. We discuss how the inhomogeneous distribution of the primary charge sequence of the CTPR proteins predicts translocation barriers that are proportional to the bias voltage. Larger barriers at higher bias voltages will result in longer translocation times, consistent with our experimental results.

3099-Pos

Quartz Nanopore Membranes for Low Noise Measurements of Ion Channel Conductance

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Planar lipid bilayer (PLB) apparatus provide an excellent platform for the study of isolated membrane proteins. The noise performance and bandwidth of PLB systems are poor relative to the state of the art in patch clamp/pipette apparatus. The problem is the relatively high capacitance of PLB systems relative to small area patch pipettes. At low frequencies (hundreds of Hz), the difference is small to non-existent. At higher frequencies, the noise becomes dominated by voltage noise from the amplifier acting on capacitance of the lipid bilayer and surrounding platform. With a much larger area, the noise for the PLB system rapidly exceeds that of the patch pipette. At intermediate frequencies (1 to 10^3 of kHz), the specific composition of the PLB platform can lead to an increase in noise due to dielectric loss [1].

We have developed a PLB system based upon a quartz nanopore membrane (QNM) with noise performance approaching the state of the art for patch clamp systems. Due to low dielectric loss, the QNM represents a significant advance in performance over the previously presented glass nanopore membrane [2] and provides for noise performance of ~ 200 fA at a 10 kHz bandwidth when coupled to a simple capacitive feedback amplifier. The resulting system has great immunity to vibration and electrical interference, without the need for a vibration isolation table and a large Faraday shield. This new PLB platform will open up the potential for making very high bandwidth single channel measurements that were not previously possible.

[1] R. A. Levis, et al, *Methods in Enzymology*, 293, pp. 218-266, 1998

[2] R. J. White, et al, *J. Am. Chem. Soc.*, 129, pp. 11766-11775, 2007

3100-Pos

Control of Salt Rejection by Surface Charge Patterning in Conical Polymer Nanopores

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Biological membrane protein channels show a variety of interesting transport properties, such as ionic and molecular selectivity. Studies on biological nanopores have shown that a pore's selectivity is due to both steric and electrostatic filtration of the ion or molecule that the pore is designed to transport. In the case of the aquaporin, the pore structure allows for the transport of water molecules at high flow rates without concurrent passage of ions. Careful preparation of an array of such salt rejecting channels would be useful in a variety of applications, in particular for desalination. To this effect, we have prepared "synthetic" salt rejecting channels from conical nanopores in polymer films. At low and moderate ionic strength, pores in polyimide and polyethylene terephthalate films are naturally cation-selective due to a native negative surface charge, and upon application of pressure, show salt rejection. The experimental data were supported by continuum modeling based on the Poisson-Nernst-Planck equations. The model also predicted that nanopores which contain a surface charge pattern consisting of a zone with positive surface charges next to a zone with negative surface charges should exhibit superior salt rejection capabilities compared to homogeneously charged pores. This improvement is due to the large potential barrier to ion transport created by the separation of cations and anions at the junction of the positively and negatively charged zones. Experimental and theoretical results are shown for both homogenous and surface charged patterned pores.

3101-Pos

Fabrication of Metallised Solid-State Nanopores Using Electrodeposition with Ionic Current Feedback

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In recent years, solid state nanopores fabricated in thin insulating membranes have been successfully employed as a new tool to detect and characterise the passage of DNA molecules. These nanopores circumvent some of the problems associated with protein channels, and offer the additional advantage of tunable pore size.

Although several experiments have clearly demonstrated that modulations of ionic current during translocation of RNA or DNA molecules can be used to discriminate between polynucleotides, a key challenge with nanopores is to find methods to slow down and control the DNA translocation. It has been proposed that the presence of a metallic probe located at the nanopore can potentially enhance the electrostatic interaction between the DNA molecule and nanopore surface and hence reduce translocation times. Moreover, by applying an electric potential to the metallic nanopore it is possible to control the charge and ultimately allow for sorting and sizing of DNA fragments.

Here we report a novel method to fabricate these metallic nanopores with apparent diameters below 20 nm using electrochemical deposition and "on-line" ionic current feedback. Starting from large nanopores (diameter 100-200 nm) milled into gold silicon nitride membranes using a focused ion beam, we electrodeposit platinum onto the gold surface, reducing the effective pore diameter. By monitoring the ion current simultaneously, the electrodeposition process can be terminated at any pre-defined value of the pore conductance in a precisely controlled and reproducible way. Our approach is applicable to single nanopores as well as nanopore arrays, and can easily be extended to metal deposits other than Pt. In order to highlight their potential for single-molecule biosensing applications, we also show electrophoretic translocation of lambda DNA in a proof-of-concept experiment.

3102-Pos

Lipid Bilayers in Nanopores to Vary their Diameter, Characterize Amyloid- β Aggregates and Monitor the Activity of Membrane-Active Enzymes

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This research introduces the concept of coating the surfaces of nanopores with supported lipid bilayers for previously inaccessible nanopore-based assays. Current methods for shrinking nanopores with nanometer precision entail the use of specialized instruments such as focused ion beams or electron beams. Furthermore, altering the surface chemistry of nanopores currently requires multiple chemical steps and typically takes longer than one day. The method presented here modifies the surface chemistry of nanopores within 90 min by deposition of desired lipids with various chemical headgroups. This work also demonstrates the use of lipids with acyl chains of different lengths to shrink the diameter of a nanopore with sub-nanometer precision. Remarkably, the surface of bilayer-coated nanopores is non-fouling and makes it possible to detect aggregates of the "notoriously sticky" peptide, amyloid- β ; the same nanopore without a bilayer clogged in every experiment. These non-fouling properties of nanopores coated with a fluid lipid bilayer made it possible to resolve single aggregates of amyloid- β and to characterize their true size distribution. Finally, this research took advantage of bilayer-coated nanopores to monitor the activity of the membrane-active enzyme, phospholipase D. Together the results presented here demonstrate that supported lipid bilayers can be used to alter the size and surface chemistry of nanopores reversibly. Moreover, bilayer-coated nanopores show promise to study membrane-active enzymes, membrane processes, as well as to perform nano-Coulter counter experiments on peptides that aggregate and adhere to surfaces such as amyloid- β .

3103-Pos

Nanopore-Based Sequence-Specific Detection of Duplex DNA for Genomic Profiling

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The ability of nucleic acids to form stable, sequence-specific complexes with foreign molecular probes has been exploited for a wide range of applications in life sciences, biotechnology, medicine, and forensics. Peptide nucleic acids (PNAs), nucleic acid analogs in which the negative sugar-phosphate backbone is replaced with a neutral peptide-like backbone, have been shown to display greater stability and sequence specificity to complementary ssDNA strands than natural DNA. This feature has been utilized in a number of applications